Norovirus

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INTRODUCTION

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Norovirus is one of the leading causes of acute gastroenteritis (AGE) outbreaks and a main cause of childhood-endemic AGE worldwide. The first outbreak was described in Norwalk, Ohio, in 1968 [1,2]. Illness due to norovirus was initially described as "winter vomiting disease" due to its seasonal predilection and preponderance of patients with vomiting as a primary symptom.

The epidemiology, virology, clinical manifestations, diagnosis, pathogenesis, and prevention of norovirus will be reviewed here. Issues related to management of acute viral gastroenteritis in adults are discussed separately. (See "Acute viral gastroenteritis in adults", section on 'Treatment'.)

EPIDEMIOLOGY

Burden of disease

— Norovirus is the most common viral cause of epidemic gastroenteritis worldwide; it is also a common cause of endemic diarrhea in community settings [3-6]. The highest frequency of norovirus infection occurs among infants less than 12 months of age [7].

Each year in the United States, norovirus causes 19 to 21 million illnesses, including 900 deaths, 103,000 hospitalizations, 460,000 emergency department visits, and 2.6 million outpatient visits [8]. In one surveillance study between 2014 and 2016 in the United States, the estimated incidence of medically attended episodes was 5.5 per 1000 person-years; the incidence was highest among children <5 years (20.4 per 1000 person-years) followed by adults ≥ 65 years (4.5 per 1000 person-years) [9].

In a meta-analysis including nearly 150,000 patients with acute gastroenteritis, the overall norovirus detection rate was 17 percent [10]. Birth cohort studies in low- and middle-income countries have demonstrated that up to 90 percent of the children experience at least one norovirus infection, and up to 70 percent experience norovirus-associated diarrhea in early childhood [6].

In most countries where rotavirus vaccines have been included in the national immunization program, norovirus has become the most common cause of gastroenteritis in children <5 years of age [4,11-15].

Seasonality

— Norovirus infection can be acquired at any time of year; some studies in temperate climates have noted a peak in incidence during the winter months [16,17]. As an example, in a study of over 1600 stool specimens collected from patients presenting with gastroenteritis to Veterans Affairs Medical Centers in the United States, 6 percent overall were positive for norovirus, with a higher medical prevalence of norovirus from November to April compared with the rest of the year (9 versus 3 percent) [18].

Transmission

• **Person-to-person** – Person-to-person transmission of norovirus occurs via the fecal-oral route, with an incubation period of 24 to 48 hours [19]. A small inoculum (<100 viral particles) is required for transmission [20-22].

Norovirus shedding in stool is maximal over the first 24 to 48 hours after illness; the mean duration of viral shedding is four weeks after onset of illness [23-30]. In immunocompromised hosts, viral shedding in stool can persist for months following infection [24,31].

Norovirus transmission occurs more frequently among symptomatic patients than asymptomatic shedders, and nosocomial transmission is common [32,33].

• Environmental transmission – Spread of norovirus infection can occur via airborne droplets of vomitus containing viral particles or fomite contamination [20,34-42]. Norovirus is extremely stable in the environment; it resists freezing temperatures, heating to 60°C, and disinfection with chlorine or alcohol [43].

Norovirus transmission can also occur by consumption of contaminated food and water. Foods linked to outbreaks have included shellfish (including oysters), leafy greens, produce items (such as celery, melon, and raspberries), and frosting; however, specific food items can be implicated in the minority of outbreaks [44,45]. In the United States, annual summaries of foodborne outbreaks are reported by the United States Centers for Disease Control and Prevention (CDC). (See "Causes of acute infectious diarrhea and other foodborne illnesses in resource-abundant settings".)

Outbreaks

- Norovirus outbreaks have occurred in a wide range of settings, including [1,16,46-56]:

- Restaurants and catered events
- Hospitals and long-term care facilities
- Schools, childcare settings, and community centers
- Municipal water contamination and recreational water exposure
- Cruise ships and resorts
- Military populations
- Athletic teams
- Rafters and backpackers
- Natural disasters
- Prisons

During 2021 and 2022, the 12 Norovirus Sentinel Testing and Tracking Network sites (NoroSTAT) reported 992 norovirus outbreaks to the CDC in comparison with 1056 and 343 norovirus outbreaks during the 2019 to 2020 and 2020 to 2021 surveillance years, respectively. The number of norovirus outbreaks reported by these states during prepandemic surveillance years ranged from 1219 (2015 to 2016) to 1471 (2018 to 2019). Norovirus outbreak characteristics during 2021 and 2022 were similar to those reported during prepandemic years. Most outbreaks (82 percent) were due to person-to-person spread, and the majority (59 percent) occurred in long-term care facilities. Among laboratory-confirmed outbreaks with typing information during 2021 and 2022, 43 percent were GII.4 Sydney(P16), which has been the predominant norovirus strain since its emergence during 2015 to 2016 [57].

Outbreaks of norovirus can be difficult to contain given the small inoculum required for transmission and its environmental stability.

In the United States, suspected outbreaks should be reported to public health authorities and to the CDC [3]. Features that should prompt suspicion for norovirus outbreaks are discussed below. (See 'Clinical suspicion and presumptive diagnosis' below.)

VIROLOGY

Norovirus is a genus within the Caliciviridae family of small nonenveloped ribonucleic acid (RNA) viruses, which also includes *Sapovirus*. The *Norovirus* and *Sapovirus* genera contain viruses infecting humans and other animals [1].

Noroviruses are subdivided into multiple genogroups [58-60]. Genogroups GI, GII, GIV, GVIII, and GIX include human pathogens; multiple genotypes are recognized within each genogroup [61]. Frequent recombination between strains and point mutations contribute to rapid changes in genetic diversity; many recombinants are as infectious and virulent as prototype strains [62-64]. The illnesses caused by viruses in each genogroup are clinically indistinguishable despite differences in genetic sequence, genomic recombination between strains, and receptor-binding characteristics.

The most common cause of human norovirus infection is GII (mostly GII.4, followed by other types such as GII.2 and GII.17), followed by GI and GIV. GII.4 viruses have been highly predominant and are associated with more severe outcomes than other norovirus genotypes, including higher hospitalization and death rates [48,65-70].

GII.4 strains have mutations occurring in one or more antigenic sites which increase the probability of immune escapes and emergence of new variants. Two GII.4 variants were responsible for outbreaks in Australia and New Zealand from 2005 to 2006 [71]. Subsequently, one of these strains was linked to approximately one-fourth of the outbreaks reported in the United Kingdom. In 2012, GII.4 Sydney strain (named for the location in Australia where it was first isolated) replaced GII.4 New Orleans as the predominant strain in the United States [72]. The proportion of outbreaks attributed to this new strain increased from 19 to 58 percent between September and December 2012. Prior GII infection appears to protect against subsequent infection, which has important significance for vaccine development [73]. GII.4 genotypes remain predominant in the United States and most of the world. GII.4 genotypes remain predominant in the United States and most of the world [74].

In 2014, a GII.17 variant emerged in China and has spread worldwide [75-79]. Its clinical presentation is indistinguishable from that of previously predominating strains.

PATHOGENESIS

- Immunity The role of humoral and/or cellular immunity in infection and disease prevention remains partially understood. Reinfections throughout a lifetime are extremely common; nevertheless, during childhood, symptomatic reinfection with the most common genotype (GII.4) is uncommon, suggesting that protective efficacy of a previous infection occurs against the same genotype and possibly against different genotypes within the same genogroup. However, the protective efficacy seems to be less with exposure to a norovirus that belongs to a different genogroup (and thus is less genetically similar) [80,81]. Exposure over time to a diversity of norovirus strains thus results in repeated infections, many of which will be asymptomatic [82].
- Host susceptibility Human noroviruses recognize and bind to blood group antigens in a strain-specific manner; such antigens may serve as receptors or cofactors for infection [83].

The blood group antigens recognized may vary by virus genogroup [84-87]. It has been suggested that genogroup GI viruses may bind preferentially to blood group A and O antigens, while genogroup GII viruses may bind preferentially to blood group A and B antigens [88].

However, in contrast with the above observations, a study of two norovirus outbreaks among Israeli military recruits noted no association between genogroup GII norovirus infection and ABO blood group antigens, suggesting that GII viruses are capable of infecting individuals regardless of blood type [88,89]. The ability of GII.4 genotypes to infect individuals with different secretor antigens and varying levels of expression may be a factor accounting for their predominance in the human population. Nevertheless, only a few in vivo studies support the hypothesis that different GII.4 variants have different secretor antigen affinity, and additional studies are required to clarify the ability of different secretor antigens, such as ABO, to affect susceptibility to GII.4 genotypes in a strain- or variant-dependent manner [90].

Individual norovirus strains may be capable of infecting only a subset of the human population; however, given the diverse binding profiles found among the norovirus genogroups, nearly all individuals are likely susceptible to infection. In addition, recurrent infection can occur given the diversity of norovirus strains and the lack of full cross-strain or long-term immunity.

• Intestinal physiology – Diarrhea induced by norovirus is associated with transient malabsorption of D-xylose and fat [91] and with decreased activity of brush-border enzymes including alkaline phosphatase and trehalase [92]. Absorption and brush-border

enzyme levels return to normal values within two weeks after infection.

The mechanisms of norovirus-induced vomiting and diarrhea are uncertain [93]. Gastric emptying is markedly delayed in normal adults challenged with norovirus, but the degree of delay is not correlated with the severity of vomiting, and norovirus infection has not been associated with detectable enterotoxin production [94].

Acute norovirus infection produces a reversible histopathologic lesion in the jejunum, with apparent sparing of the stomach and rectum [92,95-98]. The villi are blunted, but the mucosa is otherwise intact. Mononuclear and polymorphonuclear leukocytic infiltrations are seen in the lamina propria. On electron microscopy, the epithelial cells are intact, microvilli are shortened, and the intercellular spaces are widened.

These histopathologic changes appear within 24 hours after virus challenge (whether symptomatic or subclinical), are present at the peak of illness, and persist for a variable period of time after the illness. They generally clear within two weeks after the onset of illness, although some jejunal changes have been noted as late as six weeks after challenge.

CLINICAL MANIFESTATIONS

Norovirus infection produces a spectrum of clinical manifestations, from mild illness with fever and watery diarrhea to more severe illness with fever, vomiting, headache, and constitutional symptoms [29,80,99-102]. Asymptomatic infections are common throughout an individual's lifetime.

Symptomatic infection

Incubation period and duration

— The incubation period is generally 24 to 48 hours (range 12 to 72 hours), and onset of symptoms is typically abrupt [29]. Symptoms typically last for 48 to 72 hours with rapid recovery.

Norovirus shedding in stool is maximal over the first 24 to 48 hours after illness; the mean duration of viral shedding is four weeks after onset of illness. In immunocompromised hosts, viral shedding in stool can persist for months following infection [23-28,31,73].

Typical clinical features

- Symptoms include nausea and vomiting (nonbloody, nonbilious), watery diarrhea (nonbloody), and abdominal pain.

Vomiting is more prominent in the setting of norovirus infection than in gastroenteritis caused by most other viruses [1]. If diarrhea is present, it is generally moderate (approximately four to eight stools over a period of 24 hours). Stools lack mucous, and fecal leukocytes are not seen.

Generalized myalgias, malaise, and headache are prominent. Fever occurs in approximately half of cases. In general, patients are uncomfortable but usually do not appear severely ill, although severe dehydration can occur.

In one study including 224 children in Chile with norovirus infection, nearly 90 percent had vomiting and 60 percent had fever; watery diarrhea lasted 5 to 7 days [103]. In another study including 1637 children acute gastroenteritis in Chile, Brazil, Thailand, and the Philippines, norovirus was observed in 23.8 percent of outpatients and 17.9 percent of hospitalized patients; vomiting was more common among patients with gastroenteritis due to norovirus (88 versus 57 percent), while fever was less common (61 versus 76 percent) [15].

The white blood cell count is generally normal or may be slightly elevated; relative lymphopenia may be observed at the height of the illness. Kidney function is generally normal unless dehydration ensues.

Norovirus infection cannot easily be distinguished clinically from other causes of acute gastroenteritis, in particular rotavirus. (See 'Differential diagnosis' below.)

Severe disease and complications

— Severe manifestations have been observed among older adults, children <12 months, and among immunocompromised patients [46,104-108]. In patients with severe disease, fever occurs more commonly and illness may last several days longer than in healthy individuals.

Neurologic complications have been reported in children. In infants, norovirus infection has been associated with benign convulsions [109]. In one study from Taiwan, 20 percent of 250 pediatric norovirus infections were associated with seizure [110]. Encephalitis has been sporadically reported, mainly in Japanese children with norovirus acute gastroenteritis, and carries a poor prognosis [111,112].

Chronic sequelae

— The most commonly reported chronic sequela of norovirus infection is chronic diarrhea among immunocompromised individuals, which can last for many months, leading to wasting or failure to thrive [109]. Individuals with leukemia, lymphoma, solid organ or hematopoietic cell transplantation, or graft-versus-host disease may have profuse and prolonged watery diarrhea [113-115]. Among transplant patients, norovirus infection has been associated with histopathologic changes such as disorganization and flattening of the intestinal epithelium [116]. Stool shedding may occur for several months among immunocompromised individuals [117-119].

Other postinfectious sequelae may include dyspepsia, constipation, and/or reflux [120].

Asymptomatic infection

— Stool shedding of norovirus infection in asymptomatic individuals is common, especially in children [1,121,122]. In one metaanalysis including 81 studies, the overall prevalence of asymptomatic norovirus shedding was 7 percent and was higher among children than adults (8 versus 4 percent) [122].

Asymptomatic shedding of norovirus has diagnostic implications, since diarrhea due to another cause in an asymptomatic carrier may be misattributed to norovirus infection. In addition, asymptomatic shedding has epidemiologic implications; as an example, asymptomatic food handlers can potentially transmit infection to others, as viral loads on the hands of asymptomatic and symptomatic food handlers during outbreaks are similar [123].

DIAGNOSIS

Clinical suspicion and presumptive diagnosis

- Clinical suspicion The possibility of norovirus infection should be suspected in all patients with acute onset of vomiting and/or watery diarrhea, especially if they reside in middle- or high-income countries where rotavirus vaccines are routinely used. The diagnosis of norovirus is usually presumptive in such patients; the likelihood of norovirus is higher in the setting of an outbreak or during the winter months in temperate regions.
- Approach to laboratory testing Confirming the diagnosis with stool testing is generally not necessary, although it may be useful in immunocompromised patients with severe or persistent symptoms; identification of norovirus as the cause of symptoms could help inform discontinuation of therapies for other pathogens. Identifying the etiology can also be helpful for public health purposes during outbreaks of gastroenteritis. Multi-pathogen molecular tests for gastrointestinal pathogens are becoming more widely available and used in patients with acute diarrhea, and norovirus can be identified on these tests. (See 'Laboratory tools' below.)

In a patient with watery diarrhea with a stool molecular test positive for only norovirus, the diagnosis of norovirus acute gastroenteritis is likely. However, because of the frequency of asymptomatic norovirus shedding, molecular diagnosis of norovirus does not necessarily confirm that the symptoms are due to norovirus, particularly if other pathogens are also identified on testing. In patients with a positive norovirus test but atypical symptoms, such as dysentery/bloody diarrhea or voluminous watery stools, other pathogens (eg, bacterial causes of diarrhea, including cholera in endemic regions) should be ruled out before attributing these clinical features to norovirus. (See "Approach to the adult with acute diarrhea in resource-abundant settings", section on 'Evaluation'.)

• When to suspect an outbreak – Two or more similar illnesses resulting from a common exposure should raise suspicion for a norovirus outbreak and prompt involvement of public health officials for laboratory testing and epidemiologic investigation [124]. Criteria that should raise suspicion for an outbreak caused by norovirus include prominent vomiting, lack of fever, and absence of frank blood in stools [125,126]. In one study including more than 10,000 outbreaks, these criteria were 86 percent sensitive and 92 percent specific for association with norovirus detected by reverse-transcriptase polymerase chain reaction (RT-PCR) [126]. (See 'Outbreaks' above.)

Laboratory tools

— Laboratory confirmation of norovirus is generally not necessary in clinical settings, although it may be useful in select situations (see 'Clinical suspicion and presumptive diagnosis' above).

Laboratory testing is used by public health laboratories for outbreak detection and to monitor the success of interventions to interrupt transmission.

Laboratory tools for detection of norovirus include genomic amplification via RT-PCR and antigen detection via enzyme immunoassays.

• Stool RT-PCR – Stool RT-PCR is the mainstay of laboratory diagnosis; noroviruses and other viral causes of gastroenteritis are readily detectable with this tool [127-131]. Carefully selected primers within divergent capsid regions can be used for genotypic differentiation of viral strains [132]. PCR techniques are also used widely for viral detection in food and environmental samples [84,86,133-136]. Viral detection by RT-PCR in stool may be limited by low virus concentration, improper specimen storage, inefficient viral RNA extraction, and/or presence of reverse-transcriptase inhibitors [137]. Gene amplification diagnostic tests for stool, based on rapid nucleic acid detection are highly sensitive and specific [138]. Multiplex PCR assays may screen for multiple bacterial and viral pathogens. In some cases, more than one pathogen may be detected; in these circumstances, clinical judgment is required to determine the likely causative microorganism.

Interpretation of a positive RT-PCR or immunoassay in a patient with acute diarrhea may be difficult since asymptomatic shedding is common and RT-PCR can detect a low viral load (<100 particles/gram), which may not necessarily establish norovirus as the etiology of illness. Diagnostic tests for noroviruses must be interpreted together with epidemiologic and clinical factors. This is especially relevant with the increasing use of highly sensitive, multipathogen molecular techniques, which commonly detect multiple potential pathogens in the same sample [139].

• Antigen detection – A number of antigen-detection enzyme immunoassays have been developed; these assays have lower sensitivity and specificity than RT-PCR [140-145]. Their utility is limited for diagnosis of sporadic cases of gastroenteritis, but they can be useful in outbreak settings for which multiple samples are available for testing [142,146-148].

DIFFERENTIAL DIAGNOSIS

The differential diagnosis of norovirus infection includes other causes of viral gastroenteritis as well as other causes (infectious and noninfectious) of gastrointestinal symptoms. Routine laboratory testing is not required to establish a specific etiology for acute gastroenteritis. (See "Acute viral gastroenteritis in adults", section on 'Differential diagnosis' and "Acute viral gastroenteritis in children in resource-abundant countries: Clinical features and diagnosis", section on 'Etiology'.)

Other causes of viral gastroenteritis include:

• Rotavirus – Rotavirus infection most commonly occurs in children between six months and two years of age [149,150]. In temperate northern climates, it occurs in the fall and winter; in tropical climates, and to some extent in countries in the Southern Hemisphere, it occurs throughout the year. Rotavirus infection can be difficult to distinguish from norovirus infection; moderate-to-severe gastroenteritis (characterized by vomiting in over 80 percent, watery diarrhea lasting five to seven days, and fever in over 60 percent) is common for both pathogens [103]. Dehydration and need for hospitalization are more common in rotavirus than norovirus acute gastroenteritis. (See "Clinical manifestations and diagnosis of rotavirus infection".)

- Enteric adenovirus Enteric adenoviruses (adenovirus serotypes 40 and 41) are infrequent causes of endemic gastroenteritis among children in temperate climates [151-154]. Distinguishing features include absence of vomiting, incubation period up to 10 days, and duration of symptoms up to two weeks [19,155].
- Astrovirus Astroviruses include eight serotypes (HAstV-1 to HAstV-8) and have a worldwide distribution [156-158]. Astrovirus infection occurs in individuals of all ages; it appears to be less pathogenic among adults than norovirus infection [159,160]. In temperate regions, there is a peak in infection during winter months; in tropical regions, infection occurs most frequently during rainy seasons [161,162]. Sporadic astrovirus gastroenteritis occurs primarily in children younger than four years. The incubation period is three to four days. Clinical manifestations include low-grade fever, diarrhea, headache, malaise, and nausea; vomiting occurs relatively infrequently [163-165]. Symptoms generally last two to three days.
- **Sapovirus** Sapovirus infection has a similar age distribution as norovirus and occurs year-round. It is generally detected at significantly lower levels compared with noroviruses. The incubation period is not well defined. Outbreaks of sapovirus infection can occur but are not characterized by high secondary attack rates (in contrast with norovirus infection).
- Other viruses Other viruses that have been associated with gastroenteritis include coronavirus [166,167], parechovirus (types 1 and 2 were previously referred to as the enterovirus "echovirus" types 22 and 23) [168-170], picobirnavirus [171,172], bocavirus (a parvovirus) [173,174], and Aichi virus [175,176]; it is uncertain whether all of these have a true pathogenic role in gastroenteritis. These viruses could potentially cause illnesses with clinical manifestations similar to illness caused by norovirus [177-181]. In addition, human parechovirus has been associated with severe extraintestinal disease in neonates [182]. (See "Enterovirus and parechovirus infections: Clinical features, laboratory diagnosis, treatment, and prevention", section on 'Clinical features of parechovirus infections'.)

Nonviral etiologies of vomiting or diarrhea are discussed further separately. (See "Causes of acute infectious diarrhea and other foodborne illnesses in resource-abundant settings" and "Approach to the adult with nausea and vomiting" and "Approach to the infant or child with nausea and vomiting" and "Approach to the adult with acute diarrhea in resource-abundant settings" and "Diagnostic approach to diarrhea in children in resource-abundant settings".)

TREATMENT

The management of norovirus infection is as described for other types of viral gastroenteritis; this is discussed separately. (See "Acute viral gastroenteritis in adults", section on 'Treatment' and "Acute viral gastroenteritis in children in resource-abundant countries: Management and prevention".)

PREVENTION AND CONTROL

• Health care settings – In inpatient settings, use of contact precautions is warranted for patients with vomiting and/or diarrhea [3,183-185]. (See "Infection prevention: Precautions for preventing transmission of infection", section on 'Isolation precautions'.)

Norovirus is not killed by alcohol; therefore, hand hygiene for caregivers of patients with gastroenteritis should consist of washing hands with soap and water rather than use of alcohol-based hand disinfection [186,187]. (See "Infection prevention: Precautions for preventing transmission of infection", section on 'Hand hygiene'.)

- Environmental cleaning Norovirus is not eliminated by disinfection with standard cleaning agents. Therefore, contaminated surfaces should be disinfected with bleach (5 to 25 tablespoons of household bleach per gallon of water) or other disinfectant approved by the Environmental Protection Agency [3,188]. In addition, individuals who clean clinical care areas that are heavily contaminated with stool or vomitus should wear protective equipment (ie, mask, gloves, and gown). (See "Infection prevention: General principles", section on 'Cleaning and disinfection'.)
- Additional guidance for specific patient groups:

- Health care workers Health care workers who have symptoms consistent with norovirus should be excluded from work until 48 to 72 hours after symptom resolution [3,189].
- **Children** Children should be excluded from child care centers until stools are contained in a diaper or when toilet-trained children no longer have accidents using the toilet, and when stool frequency becomes no more than two stools above that child's normal frequency, even if the stools remain loose [187].
- Food handlers Individuals with norovirus infection should not prepare food for others until at least two days after resolution of symptoms [188].

The Centers for Disease Control and Prevention (CDC) provides information regarding prevention of norovirus infection [190].

Efforts for development of effective vaccination are underway [191,192].

SOCIETY GUIDELINE LINKS

Links to society and government-sponsored guidelines from selected countries and regions around the world are provided separately. (See "Society guideline links: Acute diarrhea in adults" and "Society guideline links: Acute diarrhea in children".)

INFORMATION FOR PATIENTS

UpToDate offers two types of patient education materials, "The Basics" and "Beyond the Basics." The Basics patient education pieces are written in plain language, at the 5th to 6th grade reading level, and they answer the four or five key questions a patient might have about a given condition. These articles are best for patients who want a general overview and who prefer short, easy-to-read materials. Beyond the Basics patient education pieces are longer, more sophisticated, and more detailed. These articles are written at the 10th to 12th grade reading level and are best for patients who want in-depth information and are comfortable with some medical jargon.

Here are the patient education articles that are relevant to this topic. We encourage you to print or e-mail these topics to your patients. (You can also locate patient education articles on a variety of subjects by searching on "patient info" and the keyword(s) of interest.)

- Basics topics (see "Patient education: Viral gastroenteritis in adults (The Basics)" and "Patient education: Diarrhea in children (The Basics)" and "Patient education: Diarrhea in teens and adults (The Basics)" and "Patient education: Food poisoning (The Basics)")
- Beyond the Basics topics (see "Patient education: Acute diarrhea in adults (Beyond the Basics)" and "Patient education: Acute diarrhea in children (Beyond the Basics)" and "Patient education: Foodborne illness (food poisoning) (Beyond the Basics)")

SUMMARY AND RECOMMENDATIONS

- Epidemiology Norovirus is one of the leading causes of acute gastroenteritis (AGE) outbreaks worldwide and a main cause of childhood-endemic AGE. Norovirus infection can be acquired at any time of year; some studies in northern temperate climates have noted a peak in incidence during the winter months. (See 'Epidemiology' above.)
- **Transmission** Person-to-person transmission of norovirus infection occurs via the fecal-oral route, via airborne droplets of vomitus containing viral particles, fomite contamination, or consumption of contaminated food and water. Outbreaks of norovirus can be difficult to contain given the small inoculum required for transmission (<100 viral particles) and the ability of the virus to survive in the environment; it resists freezing temperatures, heating to 60°C, and disinfection with chlorine or alcohol. (See 'Transmission' above.)
- Virology The most common cause of human norovirus infection is GII (mostly GII.4 followed by GII.17 and GII.2) followed by GI and GIV. GII.4 viruses have been associated with epidemic infection and more severe outcomes than other norovirus genotypes,

including higher hospitalization and death rates. (See 'Virology' above.)

- Clinical manifestations (see 'Clinical manifestations' above):
 - Incubation period and duration The incubation period of norovirus infection is generally 24 to 48 hours (range 12 to 72 hours), and onset of symptoms is typically abrupt. Symptoms typically last for 48 to 72 hours with rapid recovery. Norovirus shedding in stool is maximal over the first 24 to 48 hours after illness; the mean duration of viral shedding (detected via polymerase chain reaction) is four weeks after onset of illness. In immunocompromised hosts, viral shedding in stool can persist for months following infection. (See 'Incubation period and duration' above.)
 - **Typical clinical features** Symptoms of norovirus infection include nausea and vomiting (nonbloody, nonbilious), watery diarrhea (nonbloody), and abdominal pain. Vomiting is more prominent in the setting of norovirus infection than in gastroenteritis caused by other viruses. If diarrhea is present, it is generally moderate. Generalized myalgias, malaise, and headache are prominent. Fever occurs in approximately half of cases. In general, patients are uncomfortable but usually do not appear severely ill, although dehydration can occur. (See 'Typical clinical features' above.)
 - Severe disease Severe manifestations have been observed among older adults, children <12 months, and among immunocompromised patients. Asymptomatic norovirus infection also occurs and can be associated with transmission. (See 'Severe disease and complications' above.)
- **Diagnosis** The possibility of norovirus infection should be suspected in patients with acute onset of vomiting and/or watery diarrhea. Norovirus is usually diagnosed presumptively in such patients. Confirming the diagnosis with stool testing is generally not necessary, although it may be useful in immunocompromised patients with severe or persistent symptoms. (See 'Diagnosis' above.)
- Outbreaks Two or more similar illnesses resulting from a common exposure should raise suspicion for a norovirus outbreak and
 prompt involvement of public health officials for laboratory testing and epidemiologic investigation. Criteria that should raise
 suspicion for an outbreak caused by norovirus include prominent vomiting, lack of fever, and absence of frank blood in stools. (See
 'Clinical suspicion and presumptive diagnosis' above and 'Outbreaks' above.)
- Management The clinical management of norovirus infection is as described for other types of viral gastroenteritis; this is discussed separately. (See "Acute viral gastroenteritis in adults", section on 'Treatment'.)
- Prevention and control Norovirus is not killed by alcohol or standard cleaning agents. Therefore, hand hygiene for caretakers of patients with gastroenteritis should consist of washing hands with soap and water (rather than use of alcohol-based hand disinfection), and contaminated environmental surfaces should be disinfected with bleach. In addition, individuals with norovirus infection should not prepare food for others until at least two days after resolution of symptoms. (See 'Prevention and control' above.)

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REFERENCES

- 1. Robilotti E, Deresinski S, Pinsky BA. Norovirus. Clin Microbiol Rev 2015; 28:134.
- 2. Bányai K, Estes MK, Martella V, Parashar UD. Viral gastroenteritis. Lancet 2018; 392:175.
- **3.** Division of Viral Diseases, National Center for Immunization and Respiratory Diseases, Centers for Disease Control and Prevention. Updated norovirus outbreak management and disease prevention guidelines. MMWR Recomm Rep 2011; 60:1.
- 4. Payne DC, Vinjé J, Szilagyi PG, et al. Norovirus and medically attended gastroenteritis in U.S. children. N Engl J Med 2013; 368:1121.

- 5. Ahmed SM, Hall AJ, Robinson AE, et al. Global prevalence of norovirus in cases of gastroenteritis: a systematic review and metaanalysis. Lancet Infect Dis 2014; 14:725.
- 6. Cannon JL, Lopman BA, Payne DC, Vinjé J. Birth Cohort Studies Assessing Norovirus Infection and Immunity in Young Children: A Review. Clin Infect Dis 2019; 69:357.
- 7. Farahmand M, Moghoofei M, Dorost A, et al. Global prevalence and genotype distribution of norovirus infection in children with gastroenteritis: A meta-analysis on 6 years of research from 2015 to 2020. Rev Med Virol 2022; 32:e2237.
- **8.** Burke RM, Mattison CP, Pindyck T, et al. Burden of Norovirus in the United States, as Estimated Based on Administrative Data: Updates for Medically Attended Illness and Mortality, 2001-2015. Clin Infect Dis 2021; 73:e1.
- 9. Burke RM, Mattison CP, Marsh Z, et al. Norovirus and Other Viral Causes of Medically Attended Acute Gastroenteritis Across the Age Spectrum: Results from the Medically Attended Acute Gastroenteritis Study in the United States. Clin Infect Dis 2021; 73:e913.
- **10.** Nguyen GT, Phan K, Teng I, et al. A systematic review and meta-analysis of the prevalence of norovirus in cases of gastroenteritis in developing countries. Medicine (Baltimore) 2017; 96:e8139.
- **11.** Lorrot M, Bon F, El Hajje MJ, et al. Epidemiology and clinical features of gastroenteritis in hospitalised children: prospective survey during a 2-year period in a Parisian hospital, France. Eur J Clin Microbiol Infect Dis 2011; 30:361.
- **12.** Bresee JS, Marcus R, Venezia RA, et al. The etiology of severe acute gastroenteritis among adults visiting emergency departments in the United States. J Infect Dis 2012; 205:1374.
- **13.** Williams DJ, Edwards KM, Payne DC, et al. Decline in gastroenteritis-related triage calls after rotavirus vaccine licensure. Pediatrics 2012; 130:e872.
- 14. Arvelo W, Hall AJ, Henao O, et al. Incidence and etiology of infectious diarrhea from a facility-based surveillance system in Guatemala, 2008-2012. BMC Public Health 2019; 19:1340.
- **15.** Safadi MA, Riera-Montes M, Bravo L, et al. The burden of norovirus disease in children: a multi-country study in Chile, Brazil, Thailand and the Philippines. Int J Infect Dis 2021; 109:77.
- **16.** Fankhauser RL, Monroe SS, Noel JS, et al. Epidemiologic and molecular trends of "Norwalk-like viruses" associated with outbreaks of gastroenteritis in the United States. J Infect Dis 2002; 186:1.
- 17. Mounts AW, Ando T, Koopmans M, et al. Cold weather seasonality of gastroenteritis associated with Norwalk-like viruses. J Infect Dis 2000; 181 Suppl 2:S284.
- **18.** Grytdal S, Browne H, Collins N, et al. Trends in Incidence of Norovirus-associated Acute Gastroenteritis in 4 Veterans Affairs Medical Center Populations in the United States, 2011-2015. Clin Infect Dis 2020; 70:40.
- 19. Blacklow NR, Greenberg HB. Viral gastroenteritis. N Engl J Med 1991; 325:252.
- **20.** Bresee JS, Widdowson MA, Monroe SS, Glass RI. Foodborne viral gastroenteritis: challenges and opportunities. Clin Infect Dis 2002; 35:748.
- 21. Teunis PF, Moe CL, Liu P, et al. Norwalk virus: how infectious is it? J Med Virol 2008; 80:1468.
- 22. Atmar RL, Opekun AR, Gilger MA, et al. Determination of the 50% human infectious dose for Norwalk virus. J Infect Dis 2014; 209:1016.
- **23.** Rockx B, De Wit M, Vennema H, et al. Natural history of human calicivirus infection: a prospective cohort study. Clin Infect Dis 2002; 35:246.
- 24. Siebenga JJ, Beersma MF, Vennema H, et al. High prevalence of prolonged norovirus shedding and illness among hospitalized patients: a model for in vivo molecular evolution. J Infect Dis 2008; 198:994.
- **25.** Okhuysen PC, Jiang X, Ye L, et al. Viral shedding and fecal IgA response after Norwalk virus infection. J Infect Dis 1995; 171:566.
- 26. Chan MC, Sung JJ, Lam RK, et al. Fecal viral load and norovirus-associated gastroenteritis. Emerg Infect Dis 2006; 12:1278.
- 27. Moe CL. Preventing norovirus transmission: how should we handle food handlers? Clin Infect Dis 2009; 48:38.
- **28.** Costantini VP, Cooper EM, Hardaker HL, et al. Epidemiologic, Virologic, and Host Genetic Factors of Norovirus Outbreaks in Long-term Care Facilities. Clin Infect Dis 2016; 62:1.
- **29.** Graham DY, Jiang X, Tanaka T, et al. Norwalk virus infection of volunteers: new insights based on improved assays. J Infect Dis 1994; 170:34.

- **30.** Thornhill TS, Kalica AR, Wyatt RG, et al. Pattern of shedding of the Norwalk particle in stools during experimentally induced gastroenteritis in volunteers as determined by immune electron microscopy. J Infect Dis 1975; 132:28.
- **31.** Koo HL, DuPont HL. Noroviruses as a potential cause of protracted and lethal disease in immunocompromised patients. Clin Infect Dis 2009; 49:1069.
- **32.** Sukhrie FH, Teunis P, Vennema H, et al. Nosocomial transmission of norovirus is mainly caused by symptomatic cases. Clin Infect Dis 2012; 54:931.
- **33.** Franck KT, Nielsen RT, Holzknecht BJ, et al. Norovirus Genotypes in Hospital Settings: Differences Between Nosocomial and Community-Acquired Infections. J Infect Dis 2015; 212:881.
- 34. Musher DM, Musher BL. Contagious acute gastrointestinal infections. N Engl J Med 2004; 351:2417.
- **35.** Marks PJ, Vipond IB, Carlisle D, et al. Evidence for airborne transmission of Norwalk-like virus (NLV) in a hotel restaurant. Epidemiol Infect 2000; 124:481.
- **36.** Centers for Disease Control and Prevention (CDC). Norovirus outbreak in an elementary school--District of Columbia, February 2007. MMWR Morb Mortal Wkly Rep 2008; 56:1340.
- **37.** Said MA, Perl TM, Sears CL. Healthcare epidemiology: gastrointestinal flu: norovirus in health care and long-term care facilities. Clin Infect Dis 2008; 47:1202.
- 38. Repp KK, Keene WE. A point-source norovirus outbreak caused by exposure to fomites. J Infect Dis 2012; 205:1639.
- 39. Repp KK, Hostetler TP, Keene WE. A norovirus outbreak related to contaminated surfaces. J Infect Dis 2013; 208:295.
- **40.** Centers for Disease Control and Prevention (CDC). Norovirus outbreak associated with ill food-service workers--Michigan, January-February 2006. MMWR Morb Mortal Wkly Rep 2007; 56:1212.
- **41.** Bonifait L, Charlebois R, Vimont A, et al. Detection and quantification of airborne norovirus during outbreaks in healthcare facilities. Clin Infect Dis 2015; 61:299.
- **42.** Saupe AA, Rounds J, Sorenson A, et al. Outbreak of Norovirus Gastroenteritis Associated With Ice Cream Contaminated by Frozen Raspberries From China-Minnesota, United States, 2016. Clin Infect Dis 2021; 73:e3701.
- **43.** Keswick BH, Satterwhite TK, Johnson PC, et al. Inactivation of Norwalk virus in drinking water by chlorine. Appl Environ Microbiol 1985; 50:261.
- 44. Hall AJ, Wikswo ME, Pringle K, et al. Vital signs: foodborne norovirus outbreaks United States, 2009-2012. MMWR Morb Mortal Wkly Rep 2014; 63:491.
- **45.** Marsh Z, Shah MP, Wikswo ME, et al. Epidemiology of Foodborne Norovirus Outbreaks United States, 2009-2015. Food Saf (Tokyo) 2018; 6:58.
- **46.** Centers for Disease Control and Prevention (CDC). Outbreak of acute gastroenteritis associated with Norwalk-like viruses among British military personnel--Afghanistan, May 2002. MMWR Morb Mortal Wkly Rep 2002; 51:477.
- **47.** Zlot A, Simckes M, Vines J, et al. Norovirus outbreak associated with a natural lake used for recreation Oregon, 2014. MMWR Morb Mortal Wkly Rep 2015; 64:485.
- **48.** Widdowson MA, Cramer EH, Hadley L, et al. Outbreaks of acute gastroenteritis on cruise ships and on land: identification of a predominant circulating strain of norovirus--United States, 2002. J Infect Dis 2004; 190:27.
- **49.** Thornton SA, Sherman SS, Farkas T, et al. Gastroenteritis in US Marines during Operation Iraqi Freedom. Clin Infect Dis 2005; 40:519.
- **50.** Centers for Disease Control and Prevention (CDC). Outbreaks of gastroenteritis associated with noroviruses on cruise ships--United States, 2002. MMWR Morb Mortal Wkly Rep 2002; 51:1112.
- **51.** Green KY, Belliot G, Taylor JL, et al. A predominant role for Norwalk-like viruses as agents of epidemic gastroenteritis in Maryland nursing homes for the elderly. J Infect Dis 2002; 185:133.
- **52.** Centers for Disease Control and Prevention (CDC). Multistate outbreak of norovirus gastroenteritis among attendees at a family reunion--Grant County, West Virginia, October 2006. MMWR Morb Mortal Wkly Rep 2007; 56:673.
- 53. Freeland AL, Vaughan GH Jr, Banerjee SN. Acute Gastroenteritis on Cruise Ships United States, 2008-2014. MMWR Morb Mortal Wkly Rep 2016; 65:1.
- 54. Brennan J, Cavallo SJ, Garman K, et al. Notes from the Field: Multiple Modes of Transmission During a Thanksgiving Day Norovirus Outbreak Tennessee, 2017. MMWR Morb Mortal Wkly Rep 2018; 67:1300.

- 55. Calderwood LE, Wikswo ME, Mattison CP, et al. Norovirus Outbreaks in Long-term Care Facilities in the United States, 2009-2018: A Decade of Surveillance. Clin Infect Dis 2022; 74:113.
- **56.** Dale AP, Miko S, Calderwood LE, et al. Outbreak of Acute Gastroenteritis Among Rafters and Backpackers in the Backcountry of Grand Canyon National Park, April-June 2022. MMWR Morb Mortal Wkly Rep 2022; 71:1207.
- **57.** Kambhampati AK, Wikswo ME, Barclay L, et al. Notes from the Field: Norovirus Outbreaks Reported Through NoroSTAT 12 States, August 2012-July 2022. MMWR Morb Mortal Wkly Rep 2022; 71:1222.
- **58.** Hutson AM, Atmar RL, Estes MK. Norovirus disease: changing epidemiology and host susceptibility factors. Trends Microbiol 2004; 12:279.
- 59. Vinjé J. Advances in laboratory methods for detection and typing of norovirus. J Clin Microbiol 2015; 53:373.
- **60.** Chhabra P, de Graaf M, Parra GI, et al. Updated classification of norovirus genogroups and genotypes. J Gen Virol 2019; 100:1393.
- 61. Prasad BVV, Atmar RL, Ramani S, et al. Norovirus replication, host interactions and vaccine advances. Nat Rev Microbiol 2025.
- **62.** Hardy ME, Kramer SF, Treanor JJ, Estes MK. Human calicivirus genogroup II capsid sequence diversity revealed by analyses of the prototype Snow Mountain agent. Arch Virol 1997; 142:1469.
- **63.** Jiang X, Espul C, Zhong WM, et al. Characterization of a novel human calicivirus that may be a naturally occurring recombinant. Arch Virol 1999; 144:2377.
- 64. Matson DO. Calicivirus RNA Recombination. In: Viral Gastroenteritis: Perspectives in Medical Virology, Desselberger U, Gray J (Eds), Elsevier, Amsterdam 2003. p.555.
- **65.** Siebenga JJ, Vennema H, Zheng DP, et al. Norovirus illness is a global problem: emergence and spread of norovirus GII.4 variants, 2001-2007. J Infect Dis 2009; 200:802.
- **66.** Desai R, Hembree CD, Handel A, et al. Severe outcomes are associated with genogroup 2 genotype 4 norovirus outbreaks: a systematic literature review. Clin Infect Dis 2012; 55:189.
- 67. Lopman B, Vennema H, Kohli E, et al. Increase in viral gastroenteritis outbreaks in Europe and epidemic spread of new norovirus variant. Lancet 2004; 363:682.
- **68.** Centers for Disease Control and Prevention (CDC). Norovirus activity--United States, 2006-2007. MMWR Morb Mortal Wkly Rep 2007; 56:842.
- **69.** Hossain ME, Rahman R, Ali SI, et al. Epidemiologic and Genotypic Distribution of Noroviruses Among Children With Acute Diarrhea and Healthy Controls in a Low-income Rural Setting. Clin Infect Dis 2019; 69:505.
- 70. Lucero Y, Matson DO, Ashkenazi S, et al. Norovirus: Facts and Reflections from Past, Present, and Future. Viruses 2021; 13.
- **71.** Tu ET, Bull RA, Greening GE, et al. Epidemics of gastroenteritis during 2006 were associated with the spread of norovirus GII.4 variants 2006a and 2006b. Clin Infect Dis 2008; 46:413.
- 72. Centers for Disease Control and Prevention (CDC). Emergence of new norovirus strain GII.4 Sydney--United States, 2012. MMWR Morb Mortal Wkly Rep 2013; 62:55.
- **73.** Saito M, Goel-Apaza S, Espetia S, et al. Multiple norovirus infections in a birth cohort in a Peruvian Periurban community. Clin Infect Dis 2014; 58:483.
- 74. Chhabra P, Tully DC, Mans J, et al. Emergence of Novel Norovirus GII.4 Variant. Emerg Infect Dis 2024; 30:163.
- 75. Matsushima Y, Ishikawa M, Shimizu T, et al. Genetic analyses of GII.17 norovirus strains in diarrheal disease outbreaks from December 2014 to March 2015 in Japan reveal a novel polymerase sequence and amino acid substitutions in the capsid region. Euro Surveill 2015; 20.
- 76. de Graaf M, van Beek J, Vennema H, et al. Emergence of a novel GII.17 norovirus End of the GII.4 era? Euro Surveill 2015; 20.
- 77. Lu J, Sun L, Fang L, et al. Gastroenteritis Outbreaks Caused by Norovirus GII.17, Guangdong Province, China, 2014-2015. Emerg Infect Dis 2015; 21:1240.
- 78. Lee CC, Feng Y, Chen SY, et al. Emerging norovirus GII.17 in Taiwan. Clin Infect Dis 2015; 61:1762.
- 79. Chan MCW, Hu Y, Chen H, et al. Global Spread of Norovirus GII.17 Kawasaki 308, 2014-2016. Emerg Infect Dis 2017; 23:1359.
- **80.** Johnson PC, Mathewson JJ, DuPont HL, Greenberg HB. Multiple-challenge study of host susceptibility to Norwalk gastroenteritis in US adults. J Infect Dis 1990; 161:18.
- **81.** O'Ryan ML, Lucero Y, Prado V, et al. Symptomatic and asymptomatic rotavirus and norovirus infections during infancy in a Chilean birth cohort. Pediatr Infect Dis J 2009; 28:879.

- 82. Bucardo F. Understanding Asymptomatic Norovirus Infections. EClinicalMedicine 2018; 2-3:7.
- **83.** Kambhampati A, Payne DC, Costantini V, Lopman BA. Host Genetic Susceptibility to Enteric Viruses: A Systematic Review and Metaanalysis. Clin Infect Dis 2016; 62:11.
- **84.** Hutson AM, Atmar RL, Graham DY, Estes MK. Norwalk virus infection and disease is associated with ABO histo-blood group type. J Infect Dis 2002; 185:1335.
- **85.** Rockx BH, Vennema H, Hoebe CJ, et al. Association of histo-blood group antigens and susceptibility to norovirus infections. J Infect Dis 2005; 191:749.
- 86. Lindesmith L, Moe C, Marionneau S, et al. Human susceptibility and resistance to Norwalk virus infection. Nat Med 2003; 9:548.
- Parrino TA, Schreiber DS, Trier JS, et al. Clinical immunity in acute gastroenteritis caused by Norwalk agent. N Engl J Med 1977; 297:86.
- **88.** Huang P, Farkas T, Marionneau S, et al. Noroviruses bind to human ABO, Lewis, and secretor histo-blood group antigens: identification of 4 distinct strain-specific patterns. J Infect Dis 2003; 188:19.
- **89.** Halperin T, Vennema H, Koopmans M, et al. No association between histo-blood group antigens and susceptibility to clinical infections with genogroup II norovirus. J Infect Dis 2008; 197:63.
- 90. Nordgren J, Svensson L. Genetic Susceptibility to Human Norovirus Infection: An Update. Viruses 2019; 11.
- 91. Acute infectious nonbacterial gastroenteritis: etiology and pathogenesis. Ann Intern Med 1972; 76:993.
- **92.** Agus SG, Dolin R, Wyatt RG, et al. Acute infectious nonbacterial gastroenteritis: intestinal histopathology. Histologic and enzymatic alterations during illness produced by the Norwalk agent in man. Ann Intern Med 1973; 79:18.
- **93.** Levy AG, Widerlite L, Schwartz CJ, et al. Jejunal adenylate cyclase activity in human subjects during viral gastroenteritis. Gastroenterology 1976; 70:321.
- Meeroff JC, Schreiber DS, Trier JS, Blacklow NR. Abnormal gastric motor function in viral gastroenteritis. Ann Intern Med 1980; 92:370.
- **95.** Dolin R, Levy AG, Wyatt RG, et al. Viral gastroenteritis induced by the Hawaii agent. Jejunal histopathology and serologic response. Am J Med 1975; 59:761.
- **96.** Schreiber DS, Blacklow NR, Trier JS. The mucosal lesion of the proximal small intestine in acute infectious nonbacterial gastroenteritis. N Engl J Med 1973; 288:1318.
- **97.** Schreiber DS, Blacklow NR, Trier JS. The small intestinal lesion induced by Hawaii agent acute infectious nonbacterial gastroenteritis. J Infect Dis 1974; 129:705.
- **98.** Widerlite L, Trier JS, Blacklow NR, Schreiber DS. Structure of the gastric mucosa in acute infectious bacterial gastroenteritis. Gastroenterology 1975; 68:425.
- 99. Dolin R, Treanor JJ, Madore HP. Novel agents of viral enteritis in humans. J Infect Dis 1987; 155:365.
- 100. Estes MK, Prasad BV, Atmar RL. Noroviruses everywhere: has something changed? Curr Opin Infect Dis 2006; 19:467.
- 101. Ryder RW, Greenberg H, Singh N, et al. Seroepidemiology of heat-labile enterotoxigenic Escherichia coli and Norwalk virus infections in Panamanians, Canal Zone residents, Apache Indians, and United States Peace Corps volunteers. Infect Immun 1982; 37:903.
- **102.** Zhu S, Regev D, Watanabe M, et al. Identification of immune and viral correlates of norovirus protective immunity through comparative study of intra-cluster norovirus strains. PLoS Pathog 2013; 9:e1003592.
- **103.** O'Ryan ML, Peña A, Vergara R, et al. Prospective characterization of norovirus compared with rotavirus acute diarrhea episodes in chilean children. Pediatr Infect Dis J 2010; 29:855.
- 104. Goller JL, Dimitriadis A, Tan A, et al. Long-term features of norovirus gastroenteritis in the elderly. J Hosp Infect 2004; 58:286.
- **105.** Lopman BA, Reacher MH, Vipond IB, et al. Clinical manifestation of norovirus gastroenteritis in health care settings. Clin Infect Dis 2004; 39:318.
- **106.** Mattner F, Sohr D, Heim A, et al. Risk groups for clinical complications of norovirus infections: an outbreak investigation. Clin Microbiol Infect 2006; 12:69.
- **107.** Koo HL, Ajami NJ, Jiang ZD, et al. A nosocomial outbreak of norovirus infection masquerading as clostridium difficile infection. Clin Infect Dis 2009; 48:e75.
- 108. Bok K, Green KY. Norovirus gastroenteritis in immunocompromised patients. N Engl J Med 2012; 367:2126.

- **109.** Petrignani M, Verhoef L, de Graaf M, et al. Chronic sequelae and severe complications of norovirus infection: A systematic review of literature. J Clin Virol 2018; 105:1.
- **110.** Chen YFE, Wang CY, Chiu CH, et al. Molecular epidemiology and clinical characteristics of norovirus gastroenteritis with seizures in children in Taiwan, 2006-2015. Medicine (Baltimore) 2019; 98:e17269.
- 111. Shima T, Okumura A, Kurahashi H, et al. A nationwide survey of norovirus-associated encephalitis/encephalopathy in Japan. Brain Dev 2019; 41:263.
- 112. Sánchez-Fauquier A, González-Galán V, Arroyo S, et al. Norovirus-associated encephalitis in a previously healthy 2-year-old girl. Pediatr Infect Dis J 2015; 34:222.
- **113.** Ghosh N, Malik FA, Daver RG, et al. Viral associated diarrhea in immunocompromised and cancer patients at a large comprehensive cancer center: a 10-year retrospective study. Infect Dis (Lond) 2017; 49:113.
- 114. Echenique IA, Stosor V, Gallon L, et al. Prolonged norovirus infection after pancreas transplantation: a case report and review of chronic norovirus. Transpl Infect Dis 2016; 18:98.
- 115. Avery RK, Lonze BE, Kraus ES, et al. Severe chronic norovirus diarrheal disease in transplant recipients: Clinical features of an under-recognized syndrome. Transpl Infect Dis 2017; 19.
- **116.** Karandikar UC, Crawford SE, Ajami NJ, et al. Detection of human norovirus in intestinal biopsies from immunocompromised transplant patients. J Gen Virol 2016; 97:2291.
- 117. Mai H, Gao Y, Cong X, et al. GII.4 Sydney_2012 norovirus infection in immunocompromised patients in Beijing and its rapid evolution in vivo. J Med Virol 2016; 88:224.
- **118.** Teunis PF, Sukhrie FH, Vennema H, et al. Shedding of norovirus in symptomatic and asymptomatic infections. Epidemiol Infect 2015; 143:1710.
- **119.** Roddie C, Paul JP, Benjamin R, et al. Allogeneic hematopoietic stem cell transplantation and norovirus gastroenteritis: a previously unrecognized cause of morbidity. Clin Infect Dis 2009; 49:1061.
- **120.** Porter CK, Faix DJ, Shiau D, et al. Postinfectious gastrointestinal disorders following norovirus outbreaks. Clin Infect Dis 2012; 55:915.
- 121. Phillips G, Tam CC, Rodrigues LC, Lopman B. Prevalence and characteristics of asymptomatic norovirus infection in the community in England. Epidemiol Infect 2010; 138:1454.
- **122.** Qi R, Huang YT, Liu JW, et al. Global Prevalence of Asymptomatic Norovirus Infection: A Meta-analysis. EClinicalMedicine 2018; 2-3:50.
- 123. Sabrià A, Pintó RM, Bosch A, et al. Norovirus shedding among food and healthcare workers exposed to the virus in outbreak settings. J Clin Virol 2016; 82:119.
- 124. Centers for Disease Control and Prevention. Reporting and Surveillance for Norovirus. https://www.cdc.gov/norovirus/reporting/index.html (Accessed on March 23, 2017).
- **125.** Kaplan JE, Feldman R, Campbell DS, et al. The frequency of a Norwalk-like pattern of illness in outbreaks of acute gastroenteritis. Am J Public Health 1982; 72:1329.
- **126.** Lively JY, Johnson SD, Wikswo M, et al. Clinical and Epidemiologic Profiles for Identifying Norovirus in Acute Gastroenteritis Outbreak Investigations. Open Forum Infect Dis 2018; 5:ofy049.
- **127.** Jiang X, Wang J, Graham DY, Estes MK. Detection of Norwalk virus in stool by polymerase chain reaction. J Clin Microbiol 1992; 30:2529.
- **128.** Moe CL, Gentsch J, Ando T, et al. Application of PCR to detect Norwalk virus in fecal specimens from outbreaks of gastroenteritis. J Clin Microbiol 1994; 32:642.
- 129. Schwab KJ, Estes MK, Neill FH, Atmar RL. Use of heat release and an internal RNA standard control in reverse transcription-PCR detection of Norwalk virus from stool samples. J Clin Microbiol 1997; 35:511.
- 130. Ando T, Monroe SS, Noel JS, Glass RI. A one-tube method of reverse transcription-PCR to efficiently amplify a 3-kilobase region from the RNA polymerase gene to the poly(A) tail of small round-structured viruses (Norwalk-like viruses). J Clin Microbiol 1997; 35:570.
- **131.** Kundu S, Lockwood J, Depledge DP, et al. Next-generation whole genome sequencing identifies the direction of norovirus transmission in linked patients. Clin Infect Dis 2013; 57:407.

- **132.** Ando T, Monroe SS, Gentsch JR, et al. Detection and differentiation of antigenically distinct small round-structured viruses (Norwalk-like viruses) by reverse transcription-PCR and southern hybridization. J Clin Microbiol 1995; 33:64.
- **133.** Atmar RL, Neill FH, Romalde JL, et al. Detection of Norwalk virus and hepatitis A virus in shellfish tissues with the PCR. Appl Environ Microbiol 1995; 61:3014.
- **134.** Atmar RL, Neill FH, Woodley CM, et al. Collaborative evaluation of a method for the detection of Norwalk virus in shellfish tissues by PCR. Appl Environ Microbiol 1996; 62:254.
- **135.** Atmar RL, Metcalf TG, Neill FH, Estes MK. Detection of enteric viruses in oysters by using the polymerase chain reaction. Appl Environ Microbiol 1993; 59:631.
- 136. Beuret C. Simultaneous detection of enteric viruses by multiplex real-time RT-PCR. J Virol Methods 2004; 115:1.
- 137. Patel MM, Widdowson MA, Glass RI, et al. Systematic literature review of role of noroviruses in sporadic gastroenteritis. Emerg Infect Dis 2008; 14:1224.
- 138. McHugh MP, Guerendiain D, Hardie A, et al. Detection of Norovirus by BD MAX[™], Xpert® Norovirus, and xTAG® Gastrointestinal Pathogen Panel in stool and vomit samples. J Clin Virol 2018; 105:72.
- 139. Zhang J, Guan H, Zhao W, et al. Evaluation of the BioFire FilmArray Gastrointestinal Panel and Real-Time Polymerase Chain Reaction Assays for the Detection of Major Diarrheagenic Pathogens by a Multicenter Diarrheal Disease Surveillance Program in China. Foodborne Pathog Dis 2019; 16:788.
- 140. Marshall JA, Bruggink LD. Laboratory diagnosis of norovirus. Clin Lab 2006; 52:571.
- 141. de Bruin E, Duizer E, Vennema H, Koopmans MP. Diagnosis of Norovirus outbreaks by commercial ELISA or RT-PCR. J Virol Methods 2006; 137:259.
- 142. González GG, Liprandi F, Ludert JE. Evaluation of a commercial enzyme immunoassay for the detection of norovirus antigen in fecal samples from children with sporadic acute gastroenteritis. J Virol Methods 2006; 136:289.
- 143. Burton-MacLeod JA, Kane EM, Beard RS, et al. Evaluation and comparison of two commercial enzyme-linked immunosorbent assay kits for detection of antigenically diverse human noroviruses in stool samples. J Clin Microbiol 2004; 42:2587.
- 144. Vinjé J, Vennema H, Maunula L, et al. International collaborative study to compare reverse transcriptase PCR assays for detection and genotyping of noroviruses. J Clin Microbiol 2003; 41:1423.
- 145. Richards AF, Lopman B, Gunn A, et al. Evaluation of a commercial ELISA for detecting Norwalk-like virus antigen in faeces. J Clin Virol 2003; 26:109.
- 146. Dimitriadis A, Bruggink LD, Marshall JA. Evaluation of the Dako IDEIA norovirus EIA assay for detection of norovirus using faecal specimens from Australian gastroenteritis outbreaks. Pathology 2006; 38:157.
- 147. Dimitriadis A, Marshall JA. Evaluation of a commercial enzyme immunoassay for detection of norovirus in outbreak specimens. Eur J Clin Microbiol Infect Dis 2005; 24:615.
- 148. Herrmann JE, Nowak NA, Perron-Henry DM, et al. Diagnosis of astrovirus gastroenteritis by antigen detection with monoclonal antibodies. J Infect Dis 1990; 161:226.
- 149. Elliott EJ. Acute gastroenteritis in children. BMJ 2007; 334:35.
- **150.** Parashar UD, Nelson EA, Kang G. Diagnosis, management, and prevention of rotavirus gastroenteritis in children. BMJ 2013; 347:f7204.
- **151.** Kotloff KL, Losonsky GA, Morris JG Jr, et al. Enteric adenovirus infection and childhood diarrhea: an epidemiologic study in three clinical settings. Pediatrics 1989; 84:219.
- **152.** Uhnoo I, Wadell G, Svensson L, Johansson ME. Importance of enteric adenoviruses 40 and 41 in acute gastroenteritis in infants and young children. J Clin Microbiol 1984; 20:365.
- **153.** Cunliffe NA, Booth JA, Elliot C, et al. Healthcare-associated viral gastroenteritis among children in a large pediatric hospital, United Kingdom. Emerg Infect Dis 2010; 16:55.
- **154.** Rodriguez WJ, Kim HW, Brandt CD, et al. Fecal adenoviruses from a longitudinal study of families in metropolitan Washington, D.C.: laboratory, clinical, and epidemiologic observations. J Pediatr 1985; 107:514.
- **155.** Krajden M, Brown M, Petrasek A, Middleton PJ. Clinical features of adenovirus enteritis: a review of 127 cases. Pediatr Infect Dis J 1990; 9:636.
- **156.** Lee TW, Kurtz JB. Prevalence of human astrovirus serotypes in the Oxford region 1976-92, with evidence for two new serotypes. Epidemiol Infect 1994; 112:187.

- **157.** Noel JS, Lee TW, Kurtz JB, et al. Typing of human astroviruses from clinical isolates by enzyme immunoassay and nucleotide sequencing. J Clin Microbiol 1995; 33:797.
- **158.** Taylor MB, Walter J, Berke T, et al. Characterisation of a South African human astrovirus as type 8 by antigenic and genetic analyses. J Med Virol 2001; 64:256.
- 159. Kurtz JB, Lee TW, Craig JW, Reed SE. Astrovirus infection in volunteers. J Med Virol 1979; 3:221.
- **160.** Midthun K, Greenberg HB, Kurtz JB, et al. Characterization and seroepidemiology of a type 5 astrovirus associated with an outbreak of gastroenteritis in Marin County, California. J Clin Microbiol 1993; 31:955.
- **161.** Bates PR, Bailey AS, Wood DJ, et al. Comparative epidemiology of rotavirus, subgenus F (types 40 and 41) adenovirus and astrovirus gastroenteritis in children. J Med Virol 1993; 39:224.
- 162. Cruz JR, Bartlett AV, Herrmann JE, et al. Astrovirus-associated diarrhea among Guatemalan ambulatory rural children. J Clin Microbiol 1992; 30:1140.
- **163.** Lew JF, Moe CL, Monroe SS, et al. Astrovirus and adenovirus associated with diarrhea in children in day care settings. J Infect Dis 1991; 164:673.
- 164. Moe CL, Allen JR, Monroe SS, et al. Detection of astrovirus in pediatric stool samples by immunoassay and RNA probe. J Clin Microbiol 1991; 29:2390.
- **165.** Glass RI, Noel J, Mitchell D, et al. The changing epidemiology of astrovirus-associated gastroenteritis: a review. Arch Virol Suppl 1996; 12:287.
- 166. Caul EO, Paver WK, Clarke SK. Letter: Coronavirus particles in faeces from patients with gastroenteritis. Lancet 1975; 1:1192.
- **167.** Jevšnik M, Steyer A, Zrim T, et al. Detection of human coronaviruses in simultaneously collected stool samples and nasopharyngeal swabs from hospitalized children with acute gastroenteritis. Virol J 2013; 10:46.
- 168. Hyypiä T, Horsnell C, Maaronen M, et al. A distinct picornavirus group identified by sequence analysis. Proc Natl Acad Sci U S A 1992; 89:8847.
- **169.** Stanway G, Kalkkinen N, Roivainen M, et al. Molecular and biological characteristics of echovirus 22, a representative of a new picornavirus group. J Virol 1994; 68:8232.
- **170.** Wildenbeest JG, Benschop KS, Minnaar RP, et al. Clinical relevance of positive human parechovirus type 1 and 3 PCR in stool samples. Clin Microbiol Infect 2014; 20:O640.
- 171. Ludert JE, Hidalgo M, Gil F, Liprandi F. Identification in porcine faeces of a novel virus with a bisegmented double stranded RNA genome. Arch Virol 1991; 117:97.
- 172. Ganesh B, Bányai K, Martella V, et al. Picobirnavirus infections: viral persistence and zoonotic potential. Rev Med Virol 2012; 22:245.
- 173. Allander T, Tammi MT, Eriksson M, et al. Cloning of a human parvovirus by molecular screening of respiratory tract samples. Proc Natl Acad Sci U S A 2005; 102:12891.
- 174. Jin Y, Cheng WX, Xu ZQ, et al. High prevalence of human bocavirus 2 and its role in childhood acute gastroenteritis in China. J Clin Virol 2011; 52:251.
- 175. Yamashita T, Kobayashi S, Sakae K, et al. Isolation of cytopathic small round viruses with BS-C-1 cells from patients with gastroenteritis. J Infect Dis 1991; 164:954.
- **176.** Fourgeaud J, Lecuit MM, Pérot P, et al. Chronic Aichi Virus Infection As a Cause of Long-Lasting Multiorgan Involvement in Patients With Primary Immune Deficiencies. Clin Infect Dis 2023; 77:620.
- 177. Saikruang W, Khamrin P, Suantai B, et al. Molecular detection and characterization of Aichivirus A in adult patients with diarrhea in Thailand. J Med Virol 2014; 86:983.
- **178.** Iritani N, Kaida A, Abe N, et al. Detection and genetic characterization of human enteric viruses in oyster-associated gastroenteritis outbreaks between 2001 and 2012 in Osaka City, Japan. J Med Virol 2014; 86:2019.
- **179.** Levican J, Navas E, Orizola J, et al. Human bocavirus in children with acute gastroenteritis, Chile, 1985-2010. Emerg Infect Dis 2013; 19:1877.
- 180. Rovida F, Campanini G, Piralla A, et al. Molecular detection of gastrointestinal viral infections in hospitalized patients. Diagn Microbiol Infect Dis 2013; 77:231.
- 181. Chhabra P, Payne DC, Szilagyi PG, et al. Etiology of viral gastroenteritis in children <5 years of age in the United States, 2008-2009. J Infect Dis 2013; 208:790.</p>

- **182.** Stanway G, Joki-Korpela P, Hyypiä T. Human parechoviruses--biology and clinical significance. Rev Med Virol 2000; 10:57.
- **183.** Centers for Disease Control and Prevention (CDC). Recurring norovirus outbreaks in a long-term residential treatment facility Oregon, 2007. MMWR Morb Mortal Wkly Rep 2009; 58:694.
- **184.** MacCannell T, Umscheid CA, Agarwal RK, et al. Guideline for the prevention and control of norovirus gastroenteritis outbreaks in healthcare settings. Infect Control Hosp Epidemiol 2011; 32:939.
- 185. Barclay L, Park GW, Vega E, et al. Infection control for norovirus. Clin Microbiol Infect 2014; 20:731.
- **186.** Tuladhar E, Hazeleger WC, Koopmans M, et al. Reducing viral contamination from finger pads: handwashing is more effective than alcohol-based hand disinfectants. J Hosp Infect 2015; 90:226.
- 187. American Academy of Pediatrics. Red Book: 2018 Report of the Committee on Infectious Diseases, 31st ed, Kimberlin DW, Brady MT, Jackson MA, Long SS (Eds), American Academy of Pediatrics, Itasca, IL 2018.
- 188. Centers for Disease Control and Prevention. Preventing Norovirus Infection. https://www.cdc.gov/norovirus/preventing-infection.html (Accessed on September 25, 2017).
- 189. Centers for Disease Control and Prevention. General Information about Norovirus. https://www.cdc.gov/hai/organisms/norovirus.html (Accessed on February 17, 2017).
- 190. Centers for Disease Control and Prevention. How to Prevent Norovirus. 2025. https://www.cdc.gov/norovirus/prevention/index.html (Accessed on April 21, 2025).
- **191.** Baehner F, Bogaerts H, Goodwin R. Vaccines against norovirus: state of the art trials in children and adults. Clin Microbiol Infect 2016; 22 Suppl 5:S136.
- **192.** Leroux-Roels G, Cramer JP, Mendelman PM, et al. Safety and Immunogenicity of Different Formulations of Norovirus Vaccine Candidate in Healthy Adults: A Randomized, Controlled, Double-Blind Clinical Trial. J Infect Dis 2018; 217:597.

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